

A Benchmark set

PDB-id	Description	Length	Topology	Exploration radius	Execution time (s)	Avg. min RMSD _{NMR}	PDB-id	Description	Length	Topology	Exploration radius	Execution time (s)	Avg. min RMSD _{NMR}
2Y95	AUCG tetraloop human Xist A-repeat	15		1.3	32	0.4	1KPY	PEMV-1 P1-P2 frameshifting pseudoknot	29	P	2.6	691	1.4
1JU7	SLBP binding site	17		2.2	234	0.7	1YG3	ScYLV P1-P2 frameshifting pseudoknot	29	P	2.7	687	1.2
2KRY	Mitochondrial tRNA-MET ASL from human	17		2.6	29	0.8	2JWV	High affinity anti-NFkB RNA aptamer	30		3.4	156	0.9
2L6I	Coronaviral stemloop 2	17		0.9	114	0.3	2K63	EBSt1 of group II intron Sc.ai5y	30		4.1	173	2.0
2LP9	Pseudo-triloop from BMV	17		1.8	37	0.8	2L8H	HIV-1 TAR bound to probe	30		1.5	112	0.5
1LUU	ASL of yeast tRNA-PHE	18		1.4	176	0.4	1HW0	VS ribozyme substrate	31		9.0	1020	2.6
2JSG	Anticodon of E.coli tRNA-VAL3	18		1.1	35	0.4	1LDZ	Lead-dependent ribozyme	31		7.8	749	1.8
2LAC	ASL of b.subtilis tRNA-TYR	18		0.7	38	0.3	1NA2	P2B hairpin from human telomerase	31		2.4	755	0.9
2LBJ	ASL of b.subtilis tRNA-GLY	18		1.0	42	0.4	1MFY	C4 promoter of influenza A	32		4.1	760	1.5
2LBK	ASL of s.epidermis tRNA-GLY	18		1.5	27	0.5	1XHP	Extendd U6 ISL	33		0.8	615	0.3
1Z30	Stemloop D from BEV	19		1.3	389	0.4	2LI4	Antiterminator from Mg2+ riboswitch	33		7.7	185	2.1
2QH4	scaRNA 5' term hairpin from human telom.	19		1.8	45	0.6	1R2P	D5 from ai5y group II intron	35		6.3	896	2.3
2O32	U2 snRNA stem I from human	20		3.4	26	1.3	1R7W	D4 stem-loop B of enterovirus IRES	35		8.8	846	2.9
1BN0	SL3 hairpin	21		3.7	450	0.9	2JTP	Frameshift-inducing stem-loop in SIV	35		3.2	187	1.0
2M21	Stem IV loop of Tetrahymena telomerase	22		2.1	113	0.8	2L3E	P2a-J2a/b-P2b of human telomerase	36		3.5	478	1.1
4A4R	UAAC tetraloop	22		3.1	68	0.8	1N8X	HIV1 stem loop SL1	37		3.4	765	1.2
1PJY	HIV-1 frameshift inducing stem-loop	23		3.0	539	1.0	2FDT	Hairpin of eel LINE UhaL2	37		1.8	165	0.8
2JYM	Stemloop A of HBV HPRE	23		1.8	101	0.7	2LIV	PreQ1 riboswitch bound to preQ1	37	P	1.2	661	0.6
2K66	d3' stem of group II intron Sc.ai5y	23		1.1	91	0.3	2LUB	Helix H1 of human HAR1	38		2.3	167	1.0
1OW9	Active conf. of VS ribozyme	24		2.3	587	0.5	1MNX	Loop region of 5S rRNA	43		2.3	996	1.1
1S34	Splice site of Rous sarcoma virus	24		2.6	621	1.2	1A60	T and acceptor arm of TYMV	45	P/B	4.9	1010	2.0
3PHF	3' hairpin of TYMV pseudoknot	24		4.9	79	1.4	2L94	HIV-1 frameshift site bound to inhibitor	46		2.8	168	0.9
1LC6	U6 stem loop	25		3.6	699	1.0	1YMO	P2b-P3 p.knot from human telomerase	48	P	2.4	1140	1.0
2LK3	Yeast U2/U2 snRNA complex	25		1.2	75	0.5	2M8K	Pyr motif triple helix of k.lactis telomerase	49	P	1.9	388	0.9
2LV0	Helix-35 stemloop of e.coli 23S rRNA	25		2.9	113	0.9	2LU0	k- ζ reg. of group II intron Sc.ai5y	50		10.0	439	3.2
2QH2	CR7 term hairpin from human telom.	25		1.9	85	0.8	2KZL	GA motif of B. subtilis tyrS T box leader	56		2.4	890	1.1
1M82	cRNA promoter of influenza A	26		3.0	604	1.2	2LC8	MLV readthrough pseudoknot	57	P	5.1	347	2.3
2LSZ	A730 loop of Neurospora VS ribozyme	27		1.3	307	0.5	2M58	2'-5' AG1 lariat forming ribozyme	59	P/B	22.9	572	9.8
2LDL	HIV-1 exon splicing silencer 3	28		0.9	96	0.4							
2LJJ	Subdom. IV-B from CVB-3 IRES	28		2.7	151	1.1	Min		15		0.7	26	0.3
2LOZ	RNA claw of DNA packaging motor b29	28		2.1	125	1.0	Max		59		22.9	1140	9.8
2M4Q	Ribosometa decoding site of e.coli	28		6.0	154	1.7	Avg		30		3.4	372	1.2

Table ST1. Benchmark set details. The 60 RNA molecules represent non-redundant single-chain structures from the BMRB with more than 15 residues. **Topology (column 4):** A pseudo-knot topology is indicated with a P and bridge-topologies with B. The remaining have tree-like topologies. **Exploration radius (column 5):** The exploration radius is the largest RMSD between two NMR bundle structures and is used as a cutoff for seed-selection in the sampling procedure. **Avg min RMSD (column 6)** The min RMSD_{NMR} is the smallest C4' RMSD between any member of the KGSrna samples and a particular NMR bundle structure. The average over all NMR bundle structures is reported in column 6.

B Ensemble backbone torsional distributions

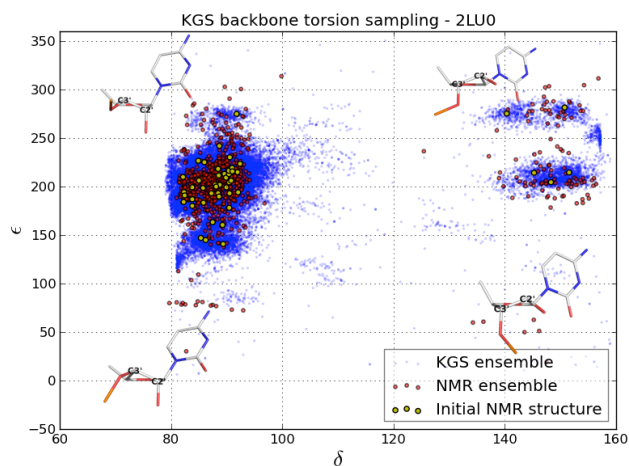


Fig. S1. Backbone $\delta - \epsilon$ torsional scatterplot of 1000 KGSrna samples of the *S. cerevisiae* group II intron (2LU0). The left cluster usually correspond to C3'-endo and the right cluster to C2'-endo ribose conformations. KGSrna extensively samples both regions as well as intermediate ones. Richardson et al. [25] suggests that ribose conformations with $\epsilon < 155^\circ$ corresponds to ribose conformations that have wrongly been fitted with C3'-endo conformations while they should have been C2'-endo. Interestingly, very few KGSrna samples lie in the region where $\epsilon < 100^\circ$.

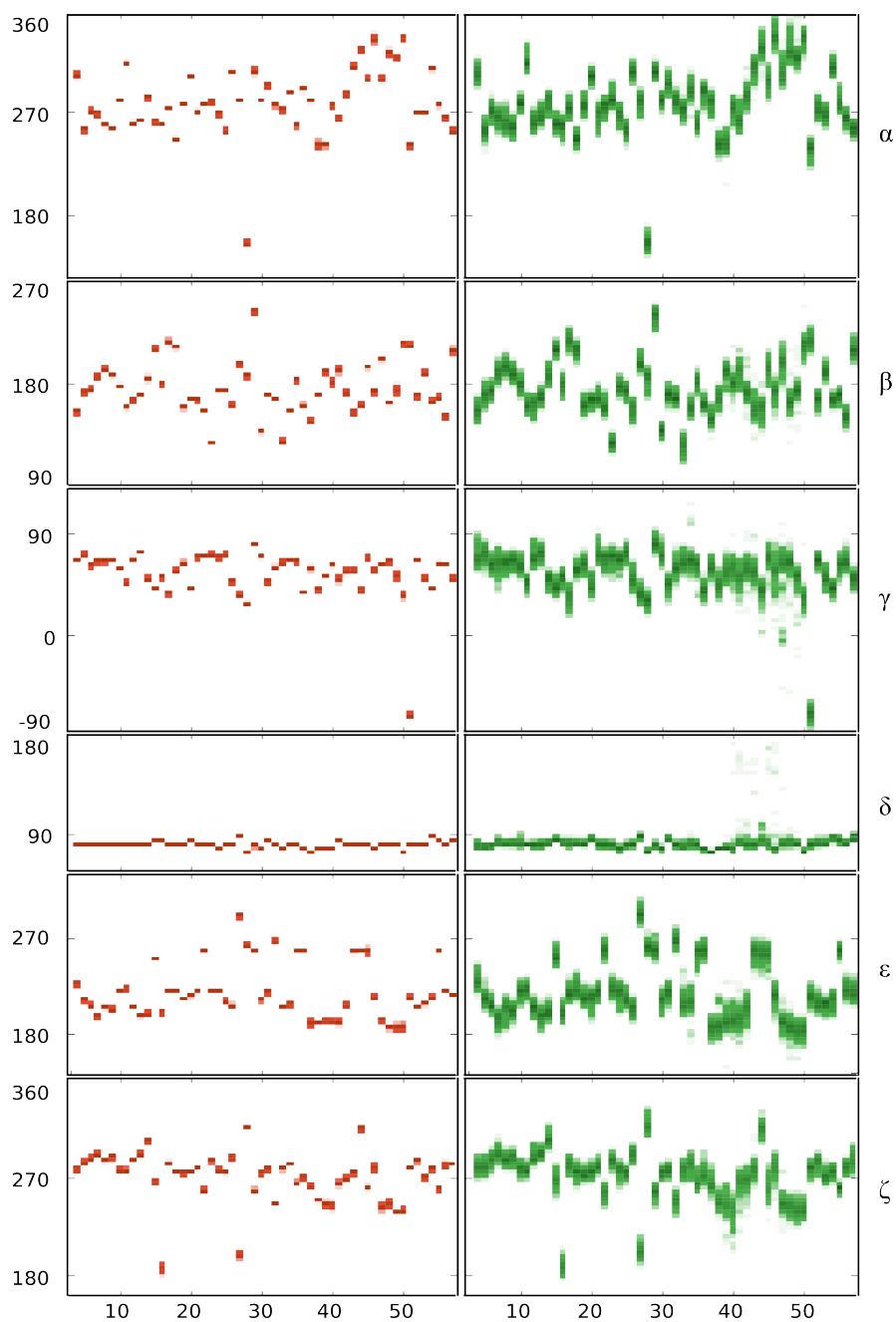


Fig. S2. Backbone torsional distributions for 1,000 iMC (left) and 1,000 KGSrna samples (right) of the MLV readthrough pseudoknot (2LC8). A wider range is sampled with KGSrna as the number of populated 3.6° bins show.

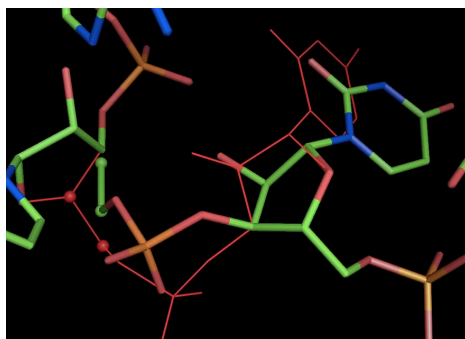
C Video files

Fig. S3. Animation showing the effect on a single nucleotide of running a rebuild perturbation. The ribose conformation has been resampled and a chain-break is introduced on the left side. The animation shows the steps as the chain is reclosed. Video available as supplementary material.

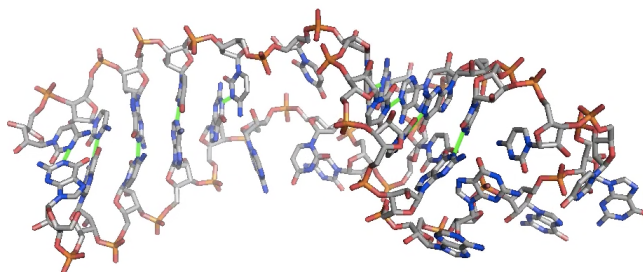


Fig. S4. Animation showing the effect on the full chain of running 100 small null space perturbations on the HIV1-TAR RNA molecule. Constraints, highlighted as green bonds, are implicitly maintained through the exact kinematics in KGSrna. Video available as supplementary material.